

REVIEW

Open Access

Blood flow-restricted exercise in space

Kyle J Hackney^{1*}, Meghan Everett^{2†}, Jessica M Scott^{3†} and Lori Ploutz-Snyder^{3†}

Abstract

Prolonged exposure to microgravity results in chronic physiological adaptations including skeletal muscle atrophy, cardiovascular deconditioning, and bone demineralization. To attenuate the negative consequences of weightlessness during spaceflight missions, crewmembers perform moderate- to high-load resistance exercise in conjunction with aerobic (cycle and treadmill) exercise. Recent evidence from ground-based studies suggests that low-load blood flow-restricted (BFR) resistance exercise training can increase skeletal muscle size, strength, and endurance when performed in a variety of ambulatory populations. This training methodology couples a remarkably low exercise training load (approximately 20%–50% one repetition maximum (1RM)) with an inflated external cuff (width, ranging between approximately 30–90 mm; pressure, ranging between approximately 100–250 mmHg) that is placed around the exercising limb. BFR aerobic (walking and cycling) exercise training methods have also recently emerged in an attempt to enhance cardiovascular endurance and functional task performance while incorporating minimal exercise intensity. Although both forms of BFR exercise training have direct implications for individuals with sarcopenia and dynapenia, the application of BFR exercise training during exposure to microgravity to prevent deconditioning remains controversial. The aim of this review is to present an overview of BFR exercise training and discuss the potential usefulness of this method as an adjunct exercise countermeasure during prolonged spaceflight. The work will specifically emphasize ambulatory BFR exercise training adaptations, mechanisms, and safety and will provide directions for future research.

Keywords: Blood flow-restricted exercise, KAATSU, Microgravity, Spaceflight, Bed rest, Unloading

Review

Introduction

Acute microgravity exposure results in rapid cephalad fluid shifts, space motion sickness, vestibular impairment, and musculoskeletal unloading [1]. However, the overall space environment encompasses both microgravity exposure and related challenges such as increased background radiation, social isolation, disruption of circadian rhythm (sunrise every 90 min), and access to a limited food variety (high salt prepackaged) and water supply [1]. These environmental stimuli interact to elicit chronic physiological adaptations including decreased bone mineral content and density, compromised maximal aerobic capacity, and reduced skeletal muscle mass and strength [2]. The morphological and structural alterations that occur within the skeletal muscle tissue as a result of microgravity exposure were uncovered following several

Shuttle Transport System, Mir, and International Space Station (ISS) investigations [3-5]. These data suggest that the rate of skeletal muscle loss relative to the duration of microgravity exposure is nonlinear, with the greatest losses early in the mission. It is also evident that skeletal muscle atrophy and dysfunction are most prominent in the knee extensor (KE; -6% to -12%) and plantar flexor (PF; -6% to -24%) muscle groups [6].

The physiological mechanisms of disuse- or unloading-related skeletal muscle atrophy have been previously reviewed [7,8]. In brief, a change in skeletal muscle size is a reflection of the temporal rates of muscle protein synthesis and degradation. For instance, across a 24-h period, if the rate of muscle protein synthesis is greater than the rate of breakdown, net muscle protein balance will be positive and protein will be gained. In contrast, if the rate of muscle protein breakdown is greater than the net rate of muscle protein synthesis, the net balance will be negative and protein will be lost. Although debated, evidence suggests that unloading induced skeletal muscle atrophy in humans occurs primarily as a result of decreased basal

* Correspondence: kyle.hackney@nasa.gov

†Equal contributors

¹Wyle Science, Technology and Engineering Group, Houston, TX 77002, USA
Full list of author information is available at the end of the article

muscle protein synthesis and a reduced synthetic response following feeding [7,9]. In-flight, variables such as participation in exercise countermeasures, age, gender, genetics, stress level, total energy intake, macronutrient composition, and preflight fitness level can influence the rates of muscle protein synthesis and breakdown; therefore, it is difficult to systematically determine the most important factor facilitating spaceflight-induced skeletal muscle atrophy, though the unloading itself is assumed to be the most potent factor.

To defend against skeletal muscle atrophy and physiological deconditioning on the ISS, crewmembers perform 2.5 h per day (including time for equipment setup and breakdown) of aerobic and/or resistance exercise; however, exercise prescription, exercise preference, and exercise adherence vary among crewmembers. The exercise equipment on the ISS, including a treadmill, a cycle ergometer, and the Advanced Resistive Exercise Device (ARED), have been engineered to operate in a weightless environment and are housed within vibration isolation systems to prevent damage to the ISS structure. During treadmill exercise, crewmembers must wear special harnesses that attach to the sides of the treadmill via bungee cords to maintain contact with the treadmill surface and to provide musculoskeletal loading during exercise. Crewmembers adjust the length of the bungee cords to set the pull-down load to a percentage (usually 60%–80%) of 1-G body mass. The cycle ergometer exercise operates similarly to an upright/recumbent bicycle and is used primarily for cardiovascular and aerobic conditioning. Resistance exercise in space is particularly challenging because the microgravity environment precludes the use of traditional free weights or weight stacks. However, ARED offers over 20 different resistance exercises including squat (SQT), heel raise, dead lift, bench press, and upright row [10]. The resistance is provided by vacuum cylinders that offer up to 273 kg of applied external load [11], and inline flywheels simulate 1-G inertia when the load changes direction during exercise. Since crewmembers are weightless in microgravity, the large loading capability is required because body mass must be added to the prescribed load in order to observe similar musculoskeletal forces to those on the ground. Thus, the current ISS exercise hardware allows for high-intensity exercise, enabling crewmembers to run up to 12 mph on the treadmill and train with heavy loads using ARED.

Although ARED and the other exercise devices provide excellent loading and resistance capabilities on the ISS, this specialized exercise hardware is expensive to build, launch, and maintain in an isolated spaceflight environment. Further, maintenance and repair must be performed by crewmembers, and spare parts for the complex devices may not be immediately available.

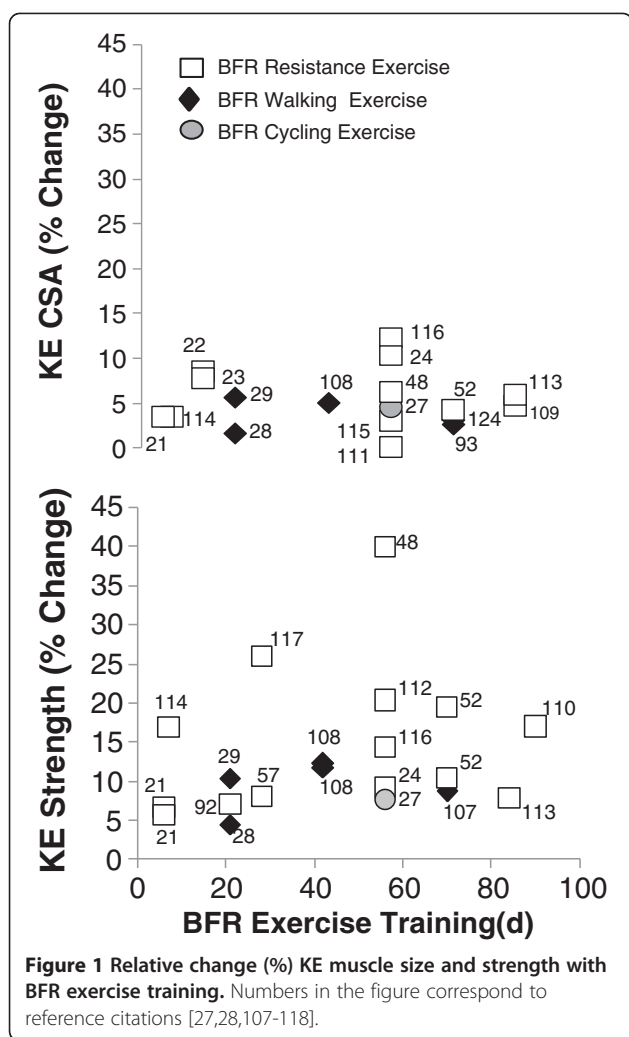
Exercise hardware for future long-duration exploration missions may need to be even more robust, compact, and portable than the current devices. There may also be fewer modes of exercise available to crewmembers, and aerobic and resistance trainings may have to be performed on a single device. It is therefore important to investigate adjunct therapies that might enhance the effectiveness of aerobic and/or resistance exercise in the event that the exercise hardware itself poses limitations. Exercising at a low intensity with blood flow restriction (via inflation cuffs) has recently become a popular method of training in Japan [12]. The purpose of this review is to present an overview of BFR exercise training and discuss its potential usefulness as an adjunct exercise countermeasure for prolonged spaceflight.

BFR exercise prescription and training

BFR resistance training

BFR resistance exercise training (also known as KAATSU when specific equipment is utilized [12]) combines low training intensity (approximately 20%–50% 1RM) with an external pressure cuff applied to the exercising limb [13–16]. BFR exercise protocols vary and are primarily influenced by cuff size, pressure, and the circumference (CIRC) of the limb being exercised [17]; however, most use three to five sets of exercise with 30- to 90-s rest periods [18]. Maintaining the cuff pressure (CP) during exercise and the rest interval also appears to be an important variable in order to increase the metabolic demand in both type I and type II muscle fibers [19]. As a result, the total number of repetitions performed in a training session can vary and is also determined by whether the first set of BFR exercise is to volitional fatigue [18] or a predetermined number of repetitions [20].

Overall, BFR exercise training studies have ranged from 6 to 90 days in duration (Figure 1) and most show dramatic muscular adaptations. For example, Fujita et al. [21] reported 6.7% and 3% increases in KE strength and size, respectively, after 6 days of twice-daily BFR knee extension (KEx). Likewise, Yasuda et al. [22] demonstrated even larger improvements in 1RM SQT (14%) and quadriceps muscle size (7.8%) from 2 weeks of twice-daily BFR SQT exercise. Moreover, BFR SQT and leg curl exercise performed 6 days per week for 2 weeks resulted in approximately 17% and 22% increases in SQT and leg curl 1RM, respectively, and an 8.5% increase in thigh cross-sectional area (CSA) [23]. Studies implementing less frequent BFR resistance exercise sessions over longer training durations also show substantial muscular adaptations. Clark et al. (2011) observed an 8% increase in KE strength after 4 weeks of BFR exercise (3 days per week), while Takarada et al. [24] reported approximately 10% increases in KE strength



and size over 8 weeks (2 days per week) of training. When studies are statistically combined, mean effect sizes ((post-mean – pre-mean)/pre-standard deviation and adjusted for sample size bias) [25] for muscle hypertrophy and strength for BFR resistance exercise are 0.39 and 0.58, respectively (compared to 0.01 and 0.000 for low-load training without cuff inflation) [15]. An important difference between high-load and BFR training is that increased muscle strength corresponds with muscle hypertrophy within the first 4 weeks of BFR exercise training, which is in contrast with the nervous system adaptations that result in enhanced muscle strength over the same duration of high-load resistance exercise training [26].

BFR aerobic training

Similar to BFR resistance training, the BFR aerobic exercise training studies couple low-intensity walking or cycling (approximately 20%–40% of maximal oxygen consumption, VO_{2max}) with an external pressure cuff of

approximately 200 mmHg applied to the upper legs [27–29]. Walk training studies showed that the metabolic cost of walking is approximately 3% higher and that heart rate (HR) is approximately 30 bpm higher during walking with BFR compared to normal walking at the same speed [29]. The training adaptations of 3-weeks of twice-daily BFR walking training at 20% of VO_{2max} included an increase in leg press and leg curl 1RM of approximately 8% and an increase in upper leg CSA of approximately 6%. No changes in muscle size or strength were observed in the control group that performed the same walking protocol without BFR [28,29]. Similarly, 3 weeks of three times-per-week BFR cycling for 15 min at 40% VO_{2max} resulted in increases in thigh CSA (3.4%), KEx strength (7.7%), and VO_{2max} (6.4%). The control group cycled for 40 min at the same intensity and showed no change in muscle strength or aerobic fitness [27]. Traditionally, aerobic exercise is prescribed at approximately 75% of VO_{2max} to elicit improvement in aerobic fitness. BFR aerobic exercise not only improves aerobic fitness at a low intensity, but also increases muscle strength and size, which are not usually observed following an aerobic training program.

Mechanisms underlying BFR exercise training adaptations

Data from BFR exercise training studies demonstrate that this novel exercise approach is gaining scientific merit as a potential alternative to traditional resistance exercise, and a growing body of literature is emerging to suggest a potential to also improve aerobic fitness. There was a recent surge in publications describing the underlying mechanisms associated with BFR resistance training adaptations; however, the physiological mechanisms explaining the observed changes remain elusive.

Muscle metabolism, motor unit recruitment, and fiber activation

Evidence suggests that type II muscle fibers are recruited at a low load during BFR exercise [19]. A current hypothesis indicates that BFR resistance exercise causes type I muscle fibers to fatigue quickly due to low oxygen availability; hence, activation of type II muscle fibers and a greater reliance on anaerobic metabolism are required [14]. These events result in an accumulation of muscle metabolites that stimulate the production of systemic or local growth factors that initiate muscle protein transcription and translation [30–32]. A convincing evidence for type II muscle fiber activation was demonstrated by Krstrup et al. [19] who showed that phosphocreatine concentrations in both slow and fast twitch fibers following BFR resistance exercise were reduced to an equivalent concentration as compared to high-intensity resistance exercise. However, fast twitch fiber recruitment evaluated by inorganic phosphate splitting

occurred in only 31% of participants performing one set of BFR resistance exercise compared to 70% of subjects performing one set of high-load exercise [32]. When multiple sets of BFR exercise were performed with continued BFR during the rest intervals, inorganic phosphate splitting was similar to multiple sets of high-load resistance exercise [33]. Therefore, the overall effort and threshold of fatigue reached during a session may facilitate the acute training response [34]. Further evidence for insufficient oxygen availability and anaerobic type II fiber activation exists in studies reporting enhanced muscle biopsy lactate [35], blood lactate (La) [20], and decreased pH [20] following BFR exercise compared to exercise at the same load without external CP [30,31].

Systemic hormonal response

Anabolic and catabolic hormonal responses have been frequently evaluated following acute and chronic BFR resistance exercise (Table 1). It is hypothesized that accumulation of metabolic by-products and/or the hypoxia-induced stimulation of afferent nerve fibers results in an increase in secretion of the growth hormone (GH) and GH-releasing hormone [36]. Takarada et al. [30] reported that circulating GH concentrations following an acute bout of BFR exercise were 290 times greater than the baseline; however, Pierce et al. [37] showed a lower but still physiologically significant ninefold increase in GH concentration using a similar protocol. A corresponding increase in the circulating insulin-like growth factor-1 (IGF1) has been observed during BFR KEx exercise (20% 1RM, four sets to exhaustion, 160–180 mmHg) and at 10–30 min post-exercise [36]. In contrast, the circulating IGF1 concentration was not increased up to 180 min following an acute

bout of BFR KEx exercise (20% 1RM, 200 mmHg, four sets, 75 total repetitions, 30-s rest periods) [20]. It is also argued that the increase in IGF1 observed in some studies could be related to a hemoconcentration as a result of plasma volume (PV) changes following BFR resistance exercise [13]. Following a training period, there was a report of a progressive increase in circulating IGF1 at rest following 2 weeks of twice-daily BFR SQT and leg curl exercise (20% 1RM, three sets, 15 repetitions/set, 30-s rest periods) [23]. Therefore, the overall relationship between BFR resistance exercise and the GH-IGF1 axis remains controversial.

The influence of acute and chronic BFR exercise on other anabolic hormones such as testosterone (T) is also unclear [38]. For example, neither Reeves et al. [39] nor Fujita et al. [20] observed changes in total or free following arm or leg BFR resistance exercise, respectively. In contrast, Madarame et al. [40] reported a post-exercise elevation in total T following three sets of BFR KEx and flexion. Chronic BFR walking [29] or resistance exercise [41] has also failed to show resting changes in T. Worthy of note, studies do show that circulating cortisol (Cort) is elevated following BFR and high-load resistance exercise [20,40,42], which suggests a similar stress response. However, circulating Cort concentration is primarily associated with catabolism and muscle protein breakdown [43]. Recently, the overall association between the acute systemic hormonal response to resistance exercise and muscle hypertrophy has been questioned. West et al. [44] reported no additional rise in muscle protein synthesis or the phosphorylation of signaling proteins following resistance exercise during elevated systemic concentrations of T, GH, and IGF1 compared to low systemic concentrations of the same anabolic hormones.

Table 1 Systemic biomarkers and hormonal responses to an acute bout of BFR exercise

Reference citation	Age (year) ^a	Exercise(s)	Intensity	Cuff width	CP (mmHg)	Significant increase ($p < 0.05$)	No change ($p > 0.05$)
[39]	21	EF,PFx	30% 1RM	NR	20↓ SBP	La, GH	T, Cort
[20]	32	KEx	20% 1RM	NR	200	La, GH, Cort	IGF1, T
[40]	26	EF,EE,KEx,KF	30% 1RM	30 × 450 mm	130–200	La, Cort, NorEpi, GH, T	
[29]	21	Walking	50 m/min, 2 min	200 × NR mm	200	GH	Cort
[30]	20–22	KEx	20% 1RM	33 × 800 mm	214	La, NorEpi, GH, IL6	CK, LP
[119]	25–40	EF	30%–50% 1RM	90 × 700 mm	0–100	La	
[120]	24–28	EF,EE,SQT,KF	20% 1RM	30 × 45 mm	190–230	La, NorEpi, GH, Hemat	Na, K,
[121]	20–22	KEx	30% 1RM	NR	200	La, NorEpi, GH	
[37]	22	KEx	20% MVC	NR	280	GH	
[21]	22	KEx	20% 1RM	NR	160–220		CK, Myo, IL6
[52]	21	EF,KEx,KF	30% 1RM	40 × 1,750 mm	>160	GH, NorEpi	T
[42]	70	KEx	20% 1RM	NR	200	La, Cort, GH	Glu
[36]	34	KEx	20% 1RM	33 × 880 mm	160–180	La, NorEpi, GH, IGF1, VEGF	Ghrl

^aAge expressed as a mean, or if not available, as a range; CP cuff pressure, NR not reported, EF elbow flexion, PFx plantar flexion, KEx knee extension, EE elbow extension, SQT squat, Myo myoglobin, La blood lactate, NorEpi norepinephrine, IL6 interleukin-6, LP lipid peroxidase, CK creatine kinase, pH blood pH, Cort cortisol, IGF1 circulating insulin-like growth factor 1, T testosterone, Na sodium, K potassium, Myo myoglobin, Glu glucose, Ghrl ghrelin, VEGF vascular endothelial growth factor.

Hence, local factors may provide greater stimuli to induce BFR resistance exercise adaptations.

Gene expression and cell signaling

An acute bout of high-load resistance exercise elicits anabolic and catabolic responses that are altered in a complex temporal manner to achieve muscle hypertrophy. Briefly, muscle protein synthesis and myogenic gene transcripts (e.g., myogenin) are upregulated within 2 h post-exercise and peak approximately 8 h post-exercise, whereas proteolytic ligases (muscle-specific RING finger protein-1 (MuRF-1), Atrogin-1) are upregulated 1–4 h after exercise and downregulated within 8 h of exercise termination [45]. An approximate threefold increase in the phosphorylation of ribosomal protein S6 kinase beta-1 (S6K1), a downstream component of the mammalian target of rapamycin (mTOR) signaling pathway and regulator of translation initiation and elongation, and a 46% increase in the fractional synthesis rate (a measure of muscle protein synthesis) have been previously shown following BFR resistance exercise [20]. mRNA expression of genes that regulate satellite cell activity (mechano-growth factor, IGF1 receptor, myogenin, MyoD), cell size (myostatin), and protein turnover (MuRF1, mTOR, S6K1) were not different between BFR and low-load exercise up to 3 h post-exercise [46]. At 8 h post-BFR or low-load exercise, myogenic gene transcripts (IGF1, MyoD, and myogenin) were not different from the baseline; however, proteolytic transcripts (Atrogin-1, MuRF-1, and Forkhead box O3 (FOXO3A)) were downregulated twofold from baseline in the BFR group only [47]. Following 8 weeks of BFR resistance training, myostatin gene expression was downregulated to a similar extent compared to high-load training [48], with a trend ($p = 0.06$) toward decreased activin IIB (myostatin receptor). There were also elevations in genes associated with myostatin function (growth and differentiation factor-associated serum protein-1) and signaling (SMAD family) [48]. Overall, the molecular pathways that regulate BFR resistance exercise-induced muscle hypertrophy have not been extensively studied. Given that muscle hypertrophy results from a positive net muscle protein balance (synthesis > breakdown) across a training period, greater examination of molecular events is needed to better understand how BFR resistance exercise stimulates muscle growth.

The limitations and safety of BFR exercise

BFR exercise limitations

The application of BFR exercise appears to be limited to peripheral muscle groups; thus, core, back, and neck muscles cannot be specifically targeted using this methodology. Higher perceptual ratings of perceived exertion and pain during the rest intervals of sets have also been reported, which could limit the application of this

training methodology [49]. The pressure applied to the blood vessel during BFR exercise is likely the root cause of discomfort and is associated with cuff width [17] and the layer of soft tissue situated between the cuff and the vessel [50]. The cuff sizes most frequently used in research studies are either narrow (approximately 5 cm) or wide (approximately 13 cm). From the data, it appears that lower cuff pressures (90–120 mmHg) are required to occlude venous blood flow when the wider cuffs are used compared to the narrow cuffs (pressures, 160–180 mmHg [51]). However, the narrow cuffs and associated pressures have repeatedly been shown to cause improvements in muscle strength and size when combined with low-load resistance exercise [52]. Exercise training adaptations have been observed with wider cuffs with the same [53,54] or lower cuff pressures [55]; however, BFR exercise performed at supra-systolic blood pressure (SBP) with wider cuffs may restrict blood flow to a level that reduces exercise volume and increases discomfort compared to narrower cuffs [56]. Therefore, cuff width is an important variable for determining BFR exercise prescription and may be a limiting factor if not taken into consideration with CP.

Hemostasis

The potential for blood coagulation and venous thrombus following blood pooling is the most frequently discussed (hypothesized) risk, and therefore, it has been comprehensively evaluated in multiple acute studies (Table 2). Most recently, Clark et al. [57] observed an increase in tissue plasminogen activator (tPA), a fibrinolytic protein that catalyzes the conversion of plasminogen to plasmin, immediately following a single bout of BFR exercise. This finding is consistent with those of Nakajima et al. [58] who reported that an acute bout of BFR exercise increased tPA antigen without altering plasminogen activator inhibitor-1 (PAI-1) or D-dimer (D-d). Together, these results suggest that BFR exercise may acutely increase fibrinolytic activity, thus reducing the risk for blood coagulation.

Acute cardiovascular stress

The acute cardiovascular responses to BFR exercise training are important to consider because this type of exercise could provide the greatest benefit for individuals with a variety of health risks that preclude them from performing traditional resistance exercise. Although most BFR literature focus on muscular effects, it is important to note that resistance and aerobic BFR exercises cause increases in HR and blood pressure that are greater than those observed with exercise performed at a similar intensity without BFR [29,59]. The increase in HR is an important aspect of BFR exercise because it allows cardiac output (CO) to be maintained, despite a

Table 2 Hemostasis markers with acute BFR resistance exercise

Reference citation	Exercise	Intensity	Sets (total repetitions)	CP (mmHg)	Cuff width (mm)	Significant increase ($p < 0.05$)	No significant change ($p > 0.05$)
[57]	KEx	30% 1RM	3 (24–36)	1.3 *×SBP	80 × 830	tPA	Fib, D-d, PTF
[20]	KEx	20% 1RM	4 (75)	200	NR		TAT, PTF 1,2, D-d
[42]	KEx	20% 1RM	4 (75)	200	NR		D-d
[122]	Leg press	30% 1RM	4 (75)	150–160	65 × 650		PTF, TAT, D-d
[58]	Leg press	30% 1RM	4 (75)	172.5	NR	tPA	D-d, Fib, PAI-1

SBP systolic blood pressure, NR not reported, TAT thrombin-anithrombin complex, tPA tissue plasminogen activator, D-d D-dimer, Fib fibrinogen, PAI-1 plasminogen activator inhibitor-1, PTF prothrombin fragment 1,2.

decrease in venous return due to the CP [36]. Since most BFR studies are conducted in healthy subjects, it is important to evaluate the cardiovascular response to BFR in individuals presenting with cardiovascular disease risk factors in a controlled setting.

Muscle damage and reperfusion

Maintaining CP during the between-set rest interval is one essential feature of the BFR exercise prescription. As a result, higher levels of muscle soreness and perceived exertion have been reported during and/or following BFR-restricted exercise compared to the same low-intensity exercise performed without CP [60,61]. Delayed onset muscle soreness is a common occurrence following eccentric muscle actions during high-load resistance exercise [62]. In contrast, it appears that concentric muscle actions compared to eccentric muscle actions result in greater muscle soreness following BFR resistance exercise [60]. It is unclear why this is the case; however, eccentric-induced muscle soreness is typically associated with mechanical stress, which may be attenuated with BFR exercise given the very low training loads. Systemic physiological markers of BFR resistance exercise-induced muscle damage are conflicting. Although neither creatine kinase nor myoglobin were elevated following two BFR resistance exercise bouts [20,30], one case of rhabdomyolysis has been reported in the literature [63].

In addition to the potential for muscle damage with BFR training, there is a hypothetical risk for microvascular dysfunction as a consequence of the reperfusion that occurs when blood flow is restored after a period of restriction or ischemia [64]. During reperfusion, there is an acute release of inflammatory molecules, clotting factors, and reactive oxidative species that impair microvascular function [65]. Further, nitric oxide bioavailability (a mediator of vasodilation) decreases when blood flow is restored, causing impaired arterial vasodilation and increased shear stress. Repeated reperfusion injury can eventually cause a wound that influences endothelial function. Renzi et al. [64] have shown a significant reduction in flow-mediated vasodilation 20 min after BFR walking exercise, suggesting the potential for endothelial dysfunction. Recent evidence also suggests increased

sarcolemma permeability (evidenced by staining of tetra-nectin) following BFR exercise, which may be caused by cell damage from increased production of reactive oxygen species [66]. Furthermore, although not statistically significant, Goldfarb et al. [67] showed that both protein carbonyls and glutathione ratios (systemic indicators of oxidative stress) were almost doubled following BFR resistance exercise in seven male subjects. Given that the time course of reactive oxygen species generation was limited to 15 min post-exercise, it is essential that future acute studies have a sufficiently high number of subjects and extended time courses.

Overall, BFR exercise training encompasses a variety of new variables (cuff width, CP, cuff inflation duration) for exercise prescription, and understanding these interactions in terms of safety is complex [68]. To date, the most comprehensive data set on side effects from BFR exercise training was established using survey methodology. The most reported incidents from approximately 13,000 people participating in KAATSU training were as follows: bruising (13.1%), numbness (1.3%), cerebral anemia (0.3%), cold feeling (0.1%), pulmonary embolism (0.01%), rhabdomyolysis (0.01%), deterioration of ischemic heart disease (0.02%), and venous thrombus (0.06%) [69].

BFR exercise in space—potential applications

Crewmembers commonly experience losses in aerobic capacity and muscular strength following long-duration spaceflight. Crewmembers that perform daily moderate-to high-intensity exercise throughout the mission duration generally return in considerably better condition than their counterparts that engage in little or low-intensity activity. Exercise with BFR may provide a means to perform resistance exercise with a low load or perform aerobic exercise at a slower walking speed or lower pedaling resistance in the event of ISS exercise hardware failure or in future exploration missions with less robust exercise hardware.

Muscle size, strength, and endurance

Cook et al. [70] recently showed that KE CSA was maintained (–1%) in a group that performed BFR resistance exercise over a 30-day period of lower-limb unloading

compared to a non-exercise control group (-7.5%). Similarly, maximal voluntary contraction (MVC) was also preserved in the BFR resistance exercise group (-2%), while the control group exhibited a 15.6% decline. Surprisingly, the BFR group also had a 28% increase in sub-maximal (approximately 40% MVC) muscular endurance compared to a 24% decline in controls. These results suggest that BFR resistance exercise may be an effective exercise countermeasure to prevent losses in KE size and strength while simultaneously improving muscular endurance [70].

The plantar flexors are another major muscle group susceptible to atrophy during unloading with the greatest plasticity evident in the soleus. For example, following 6 months on the ISS, soleus muscle size declined by 18% [71-73] and was accompanied by a decrement in PF peak torque of 20%-29% across the velocity spectrum [73]. Even more concerning is that these results were evident despite the performance of resistance and aerobic exercise countermeasures [73]. It is hypothesized that the improved loading capability (273 kg) of ARED onboard the ISS will mitigate future changes in soleus muscle size and attenuate atrophy of other muscle groups. However, the soleus muscle is best isolated when the knees are flexed, which reduces the contribution of the gastrocnemius to the muscle action [74]. Currently, this motion is difficult to reproduce using ARED because movements are primarily performed in the standing position. Another problematic aspect of targeting the soleus is a limited range of motion. As loads are increased during plantar flexion exercise, the range of motion can decrease substantially. Thus, because BFR resistance exercise is performed using low training loads, it may be possible to exercise throughout a crewmembers' full anatomic range of motion to more effectively target the soleus. Although BFR resistance exercise has not yet been performed in an unloading analog, a recent ground-based study showed a 30% increase in PF MVC following 4 weeks of training; these findings help to substantiate its potential application as a countermeasure to unloading [55].

Orthostatic challenge

BFR exercise is also a potential countermeasure to orthostatic intolerance [75-77], a condition reported to occur in up to 30% of astronauts returning from brief space shuttle flights of 4-10 days [78] and in 80% of astronauts following long-duration missions [79]. During exposure to microgravity, blood volume (BV) shifts from the capacitance vessels of the lower body to those in the face and head. Upon return to 1-G, there is excessive pooling in the lower limbs, resulting in orthostatic hypotension and syncope [80]. Research has shown that elastic cuffs worn on the upper thighs during flight help to maintain central

and peripheral hemodynamics and mitigate post-flight orthostatic intolerance [81]. However, the ability of these cuffs to completely mitigate cardiovascular deconditioning during spaceflight has not been definitively determined [82]. It is hypothesized that the most effective in-flight countermeasure would be a gravity-like stimulus, such as lower body negative pressure (LBNP). Exercise with LBNP of 1.0 to 1.2 times body weight during -6° head-down-tilt (-6HDT) bed rest has been shown to maintain upright exercise capacity [83].

BFR exercise elicits several features that are similar to LBNP including lower extremity blood pooling, decreased venous return, and increased autonomic activation [75]. Nakajima et al. [75] showed that restriction of blood flow reproduces the effects of standing on HR, stroke volume (SV), and norepinephrine release, thus simulating a gravity-like stress during short-duration -6HDT bed rest. Furthermore, Kubota et al. [76] demonstrated that BFR resistance exercise during short-duration -6HDT bed rest elicits hemodynamic and neurohumoral responses that approximate a gravity-specific stress on the cardiovascular system. In the before-mentioned study, subjects participated in 24 h of -6HDT bed rest resulting in 4.4% and 7.8% losses of BV and PV, respectively. Subjects performed BFR leg press resistance exercise (30% 1RM, four sets, 30/15/15/15 repetitions, 1-min rest intervals, 65 × 650-mm cuff width, 150-160 mmHg CP), while remaining at -6HDT. SV was significantly reduced during BFR resistance exercise and was similar to the measurement obtained when standing. These results suggest that frequent applications of BFR exercise during microgravity may provide a stimulus to the cardiovascular system that simulates 1-G, which may reduce post-flight orthostatic intolerance. Future investigations should examine the impact of BFR exercise during long-duration bed rest on the physiological responses to orthostatic stress.

BFR exercise in space—unanswered questions

To date, the majority of evidence for the application of BFR in space stems from research exploring skeletal muscle physiology during acute or chronic exercise studies. Few BFR studies have focused on the acute effects or training adaptations on cardiovascular or skeletal health. Since exposure to microgravity compromises human physiology in a variety of ways, there is a clear need to expand the scope of BFR research to include other physiological systems.

Cardiovascular health

Cardiac mass decreases to levels that are well below normal in conditions of weightlessness and simulated weightlessness. Perhonen et al. [84] demonstrated that left ventricular mass (measured by magnetic resonance imaging) decreased by 15% during prolonged supine bed

rest and by 12% after short-duration spaceflight. It appears as though disuse-induced cardiac atrophy does not influence systolic function [84]; however, one of the most important clinical consequences of cardiac atrophy may be its influence on diastolic function. Invasive studies of cardiac performance before and after 2 weeks of -6HDT bed rest have shown that there is a leftward shift in the diastolic pressure-volume curve after bed rest, resulting in a smaller left ventricular end-diastolic volume for a given filling pressure [85]. Because the effects of BFR on cardiac morphology and function have not yet been investigated, it is unclear if this form of exercise could provide the stimulus required to completely prevent cardiac deconditioning. The novelty of BFR appears to be the unique combination of venous BV pooling and restricted arterial blood inflow, resulting in a decreased SV and increased HR, while maintaining CO [36]. Consequently, in contrast to traditional aerobic exercise [86], eccentric loading of the heart and prevention of cardiac atrophy may not occur during BFR exercise.

Previous studies have established that deconditioning leads to detrimental vascular changes such as endothelial dysfunction, decreased arterial compliance, and atherosclerosis [87,88]. Hesse et al. [89] found that 13 days of bed rest impaired endothelium-dependent arterial relaxation in healthy men, while Taday and colleagues [88] reported that spaceflight significantly reduced vessel compliance [88]. Alternatively, cross-sectional studies using middle-aged and older adults have found that regular aerobic exercise improves arterial compliance [90,91]. As previously mentioned, although BFR exercise appears to acutely decrease endothelial function [64], the chronic effects of BFR exercise remain equivocal. Kim et al. [92] found that arterial compliance of the large and small arteries was not affected by 4 weeks of BFR resistance training. Conversely, Ozaki et al. [93] reported that carotid arterial compliance was improved by 10 weeks of BFR walk training in elderly subjects. Given that reduced endothelial function and arterial compliance are early markers of atherosclerosis [94], surrogate markers of cardiovascular function [95], and predictors of future cardiovascular complications [96,97], further examination of both the acute and chronic effects of BFR exercise on vascular health is needed.

Skeletal health

In addition to the cardiovascular consequences associated with prolonged spaceflight, bone health is a primary concern. Bone demineralization occurs predominately in the long bones of the lower limbs, with maximal bone loss occurring in the calcaneus and the hip [98]. Skeletal unloading in astronauts can result in losses of 1%–2% per month in bone mineral density [99,100]. On the Mir year-long

mission, bone measurements of astronauts showed a 10% reduction of the lumbar vertebrae [101]. Despite having a lower mechanical load during training, there is some evidence in ambulatory subjects that BFR resistance training may be beneficial to bone. For instance, after an acute bout of BFR resistance exercise in young men, there was a significant reduction in bone resorption as evidenced by serum N-terminal cross-linking telopeptide of type I collagen [102]. Six weeks of BFR resistance exercise training in older men also showed a 21% increase in serum bone-specific alkaline phosphatase, a marker of bone formation [102]. Although the mechanism for how low-load BFR resistance exercise improves bone parameters has not been clearly established, Loenneke et al. (2012) indicate that BFR exercise may induce physiological responses as a result of interstitial fluid flow-induced shear stress within the osteocyte membrane [103] and/or the activation of vascular endothelial growth factor via the hypoxia inducible transcription factor pathway [104]. Further work is needed to determine how BFR training could protect bone health during prolonged unloading.

Perspectives

A BFR exercise training device and Russian Braslet cuffs are currently onboard the ISS. To our knowledge, these devices have not been used with exercise by ISS crewmembers. However, BFR exercise may have already been inadvertently performed by US astronauts on Skylab. In a recent historical account, it was noted that during the first few days of the mission, astronauts had significant problems adhering to the exercise workloads prescribed on the cycle ergometer [105]. At low intensities, no issues were observed; however, at high intensities, workloads could not be reached and the astronauts described significant leg fatigue and discomfort [105]. The root cause was the combination of a padded waist belt and shoulder harness that restricted circulation to the legs as training loads increased. A solution to the problem was eventually implemented, but it is plausible that the restricted circulation these crewmembers experienced was similar to the BFR cycling methods that have been recently evaluated by researchers [27].

Overall, it is unlikely that BFR exercise will be used as a standalone exercise countermeasure onboard the ISS given the significant presence of ARED among other more traditional exercise devices. However, in future space exploration missions beyond the ISS, both aerobic and resistance BFR exercise trainings may be given some consideration depending on the vehicle capacity and the capability of onboard exercise devices. Combining low-load BFR resistance exercise with the current moderate-to high-load could help attenuate skeletal muscle atrophy without excessive loading of the shoulders, lower back, and/or joints. Specifically targeting the plantar

flexors with BFR resistance exercise training may be an alternative method to prevent soleus atrophy and dysfunction. Research also suggests that BFR exercise may simulate the cardiovascular response to standing in 1-G, which could potentially reduce orthostatic intolerance upon return to earth. However, few studies have evaluated BFR exercise using a chronic ground-based unloading analog (Table 3), and only two acute studies have used -6HDT bed rest, which is the analog that reflects the fluid shift observed during spaceflight.

Future work should evaluate the safety of exercise prescription and determine the influence of new programming variables (e.g., leg CIRC, adipose tissue thickness, CP, and cuff width) on human physiology. Once acute exercise prescription is understood, BFR exercise should be evaluated across multiple physiological systems using long duration -6HDT bed rest. Measuring cardiovascular and skeletal outcomes in addition to skeletal muscle parameters during prolonged unloading is essential to enhance our understanding of the effects of this novel

method of exercise. Furthermore, because BFR exercise training has implications for sarcopenia and other muscle wasting conditions, a mechanistic understanding of the cellular pathways resulting in muscle growth would be beneficial [106].

Conclusion

Researchers should be commended for their pioneering efforts in understanding the mechanistic and adaptive responses to BFR exercise. Both BFR resistance and aerobic exercise prescription appear to be in early stages of development relative to traditional resistance and aerobic training. Currently, low-load BFR resistance exercise increases muscle size and strength in ambulatory participants and attenuates muscle atrophy and strength loss during periods of unloading. Low-intensity BFR aerobic exercise while ambulatory enhances muscle size and strength and simultaneously increases aerobic fitness. As the science behind BFR exercise training matures in the future, it is evident that this type of

Table 3 Restriction of blood flow and BFR exercise while supine or using musculoskeletal unloading models

Reference citations	Method(s)	Highlighted outcome(s)	Limitation(s)
[75]	KAATSU (65 × 650-mm cuff, 50–100 mmHg, no muscle contractions, 10 min) following 24-h -6HDT bed rest	24 h of -6HDT bed rest resulted in ↓body mass, ↓BV, ↓PV, and ↓IVCd; 10 min of 50 mmHg, KAATSU: ↑HR, ↓SV, ↓CO, ↓IVCd, ↔Hct, ↔Hb, ↔BV, and ↔PV. Authors suggest that KAATSU reproduces the effect of a gravity-like stress during simulated weightlessness	One subject developed neurocirculatory presyncope 5 min after 100 mmHg KAATSU. There were no symptoms in the remaining seven subjects
[76]	KAATSU (65 × 650-mm cuff, 150–160 mmHg) with -6HDT leg press resistance exercise (30% 1RM, four sets, repetitions: 30/15/15/15, 1-min rest between sets) following 24-h -6HDT bed rest	24 h of -6DHT bed rest: ↓body mass, ↓BV, ↓PV, and ↑Hct. KAATSU + -6DHT leg press resistance exercise: ↑HR, ↑BP, ↓SV, and ↑CO. Authors suggest that KAATSU with leg press exercise mimics the exercise hemodynamic response to exercise in 1-G	Potential conflict of interest between the KAATSU device and the journal publishing the research study
[77]	Supine with KAATSU (60 × 605 mm, 200 mmHg, no muscle contractions vs. standing)	Supine with KAATSU: ↓SV, ↑HR, ↑TPR, and ↓CO. Authors suggest that KAATSU induced hemodynamics similar to standing	Case study: potential conflict of interest between the KAATSU device and the journal publishing the research study. Fluid shift stimuli are not introduced
[123]	Supine with KAATSU (60 × 605 mm, 50–250 mmHg, no muscle contractions vs. standing)	Supine with KAATSU: ↑FVd, ↓FBf, ↓IVCd, ↓LVd, ↓CO, ↑HR, and ↑TPR. Authors suggest that KAATSU induced hemodynamics similar to standing	Fluid shift stimuli are not introduced
[70]	BFR (60 × 830 mm, 150 mmHg) KEx resistance exercise (20% MVC, three sets, repetitions to fatigue 1.5-min rest between sets) during 30 days of unloading via ULLS	Following 30 days of ULLS: ↔KE CSA, ↑KE endurance, ↓PF CSA, ↓PF MVC, ↔IGF1, and ↔IGFBP3. Authors suggest that BFR exercise is effective in maintaining muscle size and strength and improving muscular endurance during unloading	Fluid shift stimuli are not introduced. ULLS model may not be appropriate for systemic blood markers
[124]	Restriction of blood flow (77 × 770 mm, 200 mmHg, five sets, 5-min bouts, 3-min rest between sets during 14 days of cast immobilization)	Restriction of blood flow: ↔KE MVC, ↔PF CON60, ↔leg/thigh CIRC, and ↔GH. Authors suggest that restriction of blood flow to the lower extremity prevents disuse muscular weakness	Fluid shift stimuli are not introduced. Cast immobilization model differs from spaceflight musculoskeletal unloading due to joint mobility

BFR blood flow-restricted, BV blood volume, PV plasma volume, IVCd inferior vena cava diameter, HR heart rate, SV stroke volume, CO cardiac output, Hct hematocrit, Hb hemoglobin, TPR total peripheral resistance, FVd femoral vein diameter, FBf femoral arterial blood flow, LVd left ventricle end-diastolic dimension, KE knee extensor, CSA cross-sectional area, CIRC circumference, PF plantar flexor, CON60 concentric 60° sec⁻¹, MVC maximal voluntary contraction, IGF1 circulating insulin-like growth factor, IGFBP3 circulating insulin-like growth factor binding protein-3, GH growth hormone, 1RM one repetition maximum, 6HDT -6° head-down-tilt bed rest, ULLS unilateral lower limb suspension, ↓ decreased, ↑ increased, ↔ no change.

training could be applicable as an adjunct countermeasure to combat musculoskeletal and cardiovascular dysfunctions during missions beyond low-earth orbit.

Abbreviations

1RM: One repetition maximum; -6HDT: -6° head-down-tilt; ARED: Advanced resistive exercise device; BFR: Blood flow restricted; BV: Blood volume; CIRC: Circumference; CK: Creatine kinase; CO: Cardiac output; CON60: Concentric 60° sec⁻¹; Cort: Cortisol; CP: Cuff pressure; CSA: Cross-sectional area; D-d: D-dimer; EE: Elbow extension; EF: Elbow flexion; FBF: Femoral arterial blood flow; Fib: Fibrinogen; FOXO3A: Forkhead box O3; FVd: Femoral vein diameter; Ghrl: Ghrelin; Glu: Glucose; GH: Growth hormone; Hb: Hemoglobin; Hct: Hematocrit; HR: Heart rate; IGF1: Insulin-like growth factor-1; IGFBP3: Circulating insulin-like growth factor binding protein-3; IL6: Interleukin-6; ISS: International Space Station; IVCd: Inferior vena cava diameter; K: Knee extension; KE: Knee extensor; KEx: Knee extension; La: Blood lactate; LBNP: Lower body negative pressure; LP: Lipid peroxidase; LVDd: Left ventricle end-diastolic dimension; mTOR: Mammalian target of rapamycin; MuRF-1: Muscle-specific RING finger protein-1; MVC: Maximal voluntary contraction; Myo: Myoglobin; Na: Sodium; NorEpi: Norepinephrine; NR: Not reported; PAI-1: Plasminogen activator inhibitor-1; PF: Plantar flexor; PFX: Plantar flexion; pH: Blood pH; PTf: Prothrombin fragment 1,2; PV: Plasma volume; S6K1: Ribosomal protein S6 kinase beta-1; SBP: Systolic blood pressure; SQT: Squat; SV: Stroke volume; T: Testosterone; TAT: Thrombin-anthrombin complex; tPA: Tissue plasminogen activator; TPR: Total peripheral resistance; ULLS: Unilateral lower limb suspension; VEGF: Vascular endothelial growth factor.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

KH outlined a draft of the proposal and synthesized contributions from all authors. KH, ME, JS, and LPS wrote, revised, and edited the manuscript. All authors read, edited, and approved the final version of the manuscript.

Acknowledgments

We would like to thank all members of the NASA Exercise Physiology & Countermeasures Project, Lyndon B. Johnson Space Center, Houston, Texas for their support of this work.

Author details

¹Wyle Science, Technology and Engineering Group, Houston, TX 77002, USA. ²University of Houston, Houston, TX 77002, USA. ³Universities Space Research Association, Houston, TX 77002, USA.

Received: 20 April 2012 Accepted: 5 September 2012

Published: 1 December 2012

References

1. Clement G: *Fundamentals of Space Medicine*. El Segundo: Microcosm Press; 2003.
2. Buckley JC: *Space Physiology*. New York: Oxford University Press; 2006.
3. Fitts RH, Riley DR, Widrick JJ: **Functional and structural adaptations of skeletal muscle to microgravity**. *J Exp Biol* 2001, **204**:3201–3208.
4. di Prampero PE, Narici MV: **Muscles in microgravity: from fibres to human motion**. *J Biomech* 2003, **36**:403–412.
5. Adams GR, Caiozzo VJ, Baldwin KM: **Skeletal muscle unweighting: spaceflight and ground-based models**. *J Appl Physiol* 2003, **95**:2185–2201.
6. Narici MV, de Boer MD: **Disuse of the musculo-skeletal system in space and on earth**. *Eur J Appl Physiol* 2011, **111**:403–420.
7. Phillips SM, Glover EI, Rennie MJ: **Alterations of protein turnover underlying disuse atrophy in human skeletal muscle**. *J Appl Physiol* 2009, **107**:645–654.
8. Phillips SM: **Physiologic and molecular bases of muscle hypertrophy and atrophy: impact of resistance exercise on human skeletal muscle (protein and exercise dose effects)**. *Appl Physiol Nutr Metab* 2009, **34**:403–410.
9. Glover EI, Phillips SM, Oates BR, Tang JE, Tarnopolsky MA, Selby A, Smith K, Rennie MJ: **Immobilization induces anabolic resistance in human myofibrillar protein synthesis with low and high dose amino acid infusion**. *J Physiol* 2008, **586**:6049–6061.
10. Bentley JR, Leach M, McCleary F, Smith C, Norcross J, Hagan D: *Advanced Resistive Exercise Device (ARED) Man-In-The-Loop Test (MILT) (NASA Technical Report)*. Washington, D26C: National Aeronautics and Space Administration; 2006:31.
11. Loehr JA, Lee SM, English KL, Sibonga J, Smith SM, Spiering BA, Hagan RD: **Musculoskeletal adaptations to training with the advanced resistive exercise device**. *Med Sci Sports Exerc* 2011, **43**:146–156.
12. Sato Y: **The history and future of KAATSU training**. *Int J Kaatsu Train Res* 2005, **1**:1–15.
13. Wernbom M, Augustsson J, Raastad T: **Ischemic strength training: a low-load alternative to heavy resistance exercise?** *Scand J Med Sci Sports* 2008, **18**:401–416.
14. Loenneke JP, Fahs CA, Wilson JM, Bemben MG: **Blood flow restriction: the metabolite/volume threshold theory**. *Med Hypotheses* 2011, **77**:748–752.
15. Loenneke JP, Wilson JM, Marin PJ, Zourdos MC, Bemben MG: **Low intensity blood flow restriction training: a meta-analysis**. *Eur J Appl Physiol* 2012, **112**:1849–1859.
16. Manini TM, Clark BC: **Blood flow restricted exercise and skeletal muscle health**. *Exerc Sport Sci Rev* 2009, **37**:78–85.
17. Loenneke JP, Fahs CA, Rossow LM, Sherk VD, Thiebaud RS, Abe T, Bemben DA, Bemben MG: **Effects of cuff width on arterial occlusion: implications for blood flow restricted exercise**. *Eur J Appl Physiol* 2012, **112**:2903–2912.
18. Cook SB, Clark BC, Ploutz-Snyder LL: **Effects of exercise load and blood-flow restriction on skeletal muscle function**. *Med Sci Sports Exerc* 2007, **39**:1708–1713.
19. Krustруп P, Soderlund K, Relu MU, Ferguson RA, Bangsbo J: **Heterogeneous recruitment of quadriceps muscle portions and fibre types during moderate intensity knee-extensor exercise: effect of thigh occlusion**. *Scand J Med Sci Sports* 2009, **19**:576–584.
20. Fujita S, Abe T, Drummond MJ, Cadenas JG, Dreyer HC, Sato Y, Volpi E, Rasmussen BB: **Blood flow restriction during low-intensity resistance exercise increases S6K1 phosphorylation and muscle protein synthesis**. *J Appl Physiol* 2007, **103**:903–910.
21. Fujita T, Brechue WF, Kurita K, Sato Y, Abe T: **Increased muscle volume and strength following six days of low-intensity resistance training with restricted muscle blood flow**. *Int J KAATSU Res* 2008, **4**:1–8.
22. Yasuda T, Abe T, Sato Y, Midorikawa T, Kearns CF, Inoue K, Ryushi T, Ishii N: **Muscle fiber cross sectional area is increased after two weeks of twice daily KAATSU resistance training**. *Int J Kaatsu Training Res* 2005, **1**:65–70.
23. Abe T, Midorikawa T, Sata Y, Kearns CF, Inoue K, Koizumi K, Ishii N: **Skeletal muscle size and circulating IGF-1 are increased after two weeks of twice daily "KAATSU" resistance training**. *Int J Kaatsu Training Res* 2005, **1**:6–12.
24. Takarada Y, Tsuruta T, Ishii N: **Cooperative effects of exercise and occlusive stimuli on muscular function in low-intensity resistance exercise with moderate vascular occlusion**. *Jpn J Physiol* 2004, **54**:585–592.
25. Rhea MR: **Determining the magnitude of treatment effects in strength training research through the use of the effect size**. *J Strength Cond Res* 2004, **18**:918–920.
26. Moritani T, De-Vries HA: **Neural factors versus hypertrophy in the time course of muscle strength gain**. *Am J Phys Med* 1979, **58**:115–130.
27. Abe T, Fujita S, Nakajima T, Sakamaki M, Ozaki H, Ogasawara R, Sugaya M, Kudo M, Kurano M, Yasuda T, Sato Y, Ohshima H, Mukai C, Ishii N: **Effects of low-intensity cycle training with restricted leg blood flow on thigh muscle volume and VO2max in young men**. *J Sports Sci Med* 2010, **9**:452–458.
28. Abe T, Kearns CF, Fujita S, Sakamaki M, Sato Y, Brechue WF: **Skeletal muscle size and strength are increased following walk training with restricted blood flow: implications for training duration and frequency**. *Int J KAATSU Train Res* 2009, **5**:9–15.
29. Abe T, Kearns CF, Sato Y: **Muscle size and strength are increased following walk training with restricted venous blood flow from the leg muscle, Kaatsu-walk training**. *J Appl Physiol* 2006, **100**:1460–1466.
30. Takarada Y, Nakamura Y, Aruga S, Onda T, Miyazaki S, Ishii N: **Rapid increase in plasma growth hormone after low-intensity resistance exercise with vascular occlusion**. *J Appl Physiol* 2000, **88**:61–65.
31. Yoshida T, Watari H: **Effect of circulatory occlusion on human muscle metabolism during exercise and recovery**. *Eur J Appl Physiol Occup Physiol* 1997, **75**:200–205.

32. Suga T, Okita K, Morita N, Yokota T, Hirabayashi K, Horiuchi M, Takada S, Takahashi T, Omokawa M, Kinugawa S, Tsutsui H: **Intramuscular metabolism during low-intensity resistance exercise with blood flow restriction.** *J Appl Physiol* 2009, **106**:1119–1124.
33. Suga T, Okita K, Takada S, Omokawa M, Kadoguchi T, Yokota T, Hirabayashi K, Takahashi M, Morita N, Horiuchi M, Kinugawa S, Tsutsui H: **Effect of multiple set on intramuscular metabolic stress during low-intensity resistance exercise with blood flow restriction.** *Eur J Appl Physiol* 2012, doi:10.1007/s00421-012-2377-x.
34. Burd NA, Mitchell CJ, Churchward-Venne TA, Phillips SM: **Bigger weights may not beget bigger muscles: evidence from acute muscle protein synthetic responses after resistance exercise.** *Appl Physiol Nutr Metab* 2012, **37**:551–554.
35. Lundberg G, Olofsson P, Ungerstedt U, Jansson E, Sundberg CJ: **Lactate concentrations in human skeletal muscle biopsy, microdialysate and venous blood during dynamic exercise under blood flow restriction.** *Pflugers Arch* 2002, **443**:458–465.
36. Takano H, Morita T, Iida H, Asada K, Kato M, Uno K, Hirose K, Matsumoto A, Takenaka K, Hirata Y, Eto F, Nagai R, Sato Y, Nakajima T: **Hemodynamic and hormonal responses to a short-term low-intensity resistance exercise with the reduction of muscle blood flow.** *Eur J Appl Physiol* 2005, **95**:65–73.
37. Pierce JR, Clark BC, Ploutz-Snyder LL, Kanaley JA: **Growth hormone and muscle function responses to skeletal muscle ischemia.** *J Appl Physiol* 2006, **101**:1588–1595.
38. Loenneke JP, Wilson JM, Pujol TJ, Bemben MG: **Acute and chronic testosterone response to blood flow restricted exercise.** *Horm Metab Res* 2011, **43**:669–673.
39. Reeves GV, Kraemer RR, Hollander DB, Clavier J, Thomas C, Francois M, Castracane VD: **Comparison of hormone responses following light resistance exercise with partial vascular occlusion and moderately difficult resistance exercise without occlusion.** *J Appl Physiol* 2006, **101**:1616–1622.
40. Madarama H, Sasaki K, Ishii N: **Endocrine responses to upper- and lower-limb resistance exercises with blood flow restriction.** *Acta Physiol Hung* 2010, **97**:192–200.
41. Yasuda T, Fujita S, Ogasawara R, Sato Y, Abe T: **Effects of low-intensity bench press training with restricted arm muscle blood flow on chest muscle hypertrophy: a pilot study.** *Clin Physiol Funct Imaging* 2010, **30**:338–343.
42. Fry CS, Glynn EL, Drummond MJ, Timmerman KL, Fujita S, Abe T, Dhanani S, Volpi E, Rasmussen BB: **Blood flow restriction exercise stimulates mTORC1 signaling and muscle protein synthesis in older men.** *J Appl Physiol* 2010, **108**:1199–1209.
43. Kraemer WJ, Ratamess NA: **Hormonal responses and adaptations to resistance exercise and training.** *Sports Med* 2005, **35**:339–361.
44. West DW, Kujbida GW, Moore DR, Atherton P, Burd NA, Padzik JP, De Lizio M, Tang JE, Parise G, Rennie MJ, Baker SK, Phillips SM: **Resistance exercise-induced increases in putative anabolic hormones do not enhance muscle protein synthesis or intracellular signalling in young men.** *J Physiol* 2009, **587**:5239–5247.
45. Yang Y, Creer A, Jemiolo B, Trappe S: **Time course of myogenic and metabolic gene expression in response to acute exercise in human skeletal muscle.** *J Appl Physiol* 2005, **98**:1745–1752.
46. Drummond MJ, Fujita S, Abe T, Dreyer HC, Volpi E, Rasmussen BB: **Human muscle gene expression following resistance exercise and blood flow restriction.** *Medicine and science in sports and exercise* 2008, **40**:691–698.
47. Manini TM, Vincent KR, Leeuwenburgh CL, Lees HA, Kavazis AN, Borst SE, Clark BC: **Myogenic and proteolytic mRNA expression following blood flow restricted exercise.** *Acta Physiol (Oxf)* 2011, **201**:255–263.
48. Laurentino GC, Ugrinowitsch C, Roschel H, Aoki MS, Soares AG, Neves M Jr, Aihara AY, Darcf A, Tricoli V: **Strength training with blood flow restriction diminishes myostatin gene expression.** *Med Sci Sports Exerc* 2012, **44**:406–412.
49. Loenneke JP, Balapur A, Thrower AD, Barnes JT, Pujol TJ: **The perceptual responses to occluded exercise.** *Int J Sports Med* 2011, **32**:181–184.
50. Shaw JA, Murray DG: **The relationship between tourniquet pressure and underlying soft-tissue pressure in the thigh.** *J Bone Joint Surg Am* 1982, **64**:1148–1152.
51. Crenshaw AG, Hargens AR, Gershuni DH, Rydevik B: **Wide tourniquet cuffs more effective at lower inflation pressures.** *Acta Orthop Scand* 1988, **59**:447–451.
52. Madarama H, Neya M, Ochi E, Nakazato K, Sato Y, Ishii N: **Cross-transfer effects of resistance training with blood flow restriction.** *Med Sci Sports Exerc* 2008, **40**:258–263.
53. Evans C, Vance S, Brown M: **Short-term resistance training with blood flow restriction enhances microvascular filtration capacity of human calf muscles.** *J Sports Sci* 2010, **28**:999–1007.
54. Laurentino G, Ugrinowitsch C, Aihara AY, Fernandes AR, Parcell AC, Ricard M, Tricoli V: **Effects of strength training and vascular occlusion.** *Int J Sports Med* 2008, **29**:664–667.
55. Patterson SD, Ferguson RA: **Increase in calf post-occlusive blood flow and strength following short-term resistance exercise training with blood flow restriction in young women.** *Eur J Appl Physiol* 2010, **108**:1025–1033.
56. Rossow LM, Fahs CA, Loenneke JP, Thiebaud RS, Sherck VD, Bemben MG: **Cardiovascular and perceptual responses to blood-flow-restricted resistance exercise with differing restrictive cuffs.** *Clin Physiol Funct Imaging* 2012, **32**:331–337.
57. Clark BC, Manini TM, Hoffman RL, Williams PS, Guiler MK, Knutson MJ, McGlynn ML, Kushnick MR: **Relative safety of 4 weeks of blood flow-restricted resistance exercise in young, healthy adults.** *Scand J Med Sci Sports* 2011, **21**:653–662.
58. Nakajima T, Takano H, Kurano M, Lida H, Kubota N, Yasuda T, Kato Y, Meguro K, Sato Y, Yamazaki Y, Kawashima S, Ohshima H, Tachibana S, Nagata T, Abe T, Ishii N, Morita T: **Effects of KAATSU training on haemostasis in healthy subjects.** *Int J Kaatsu Train Res* 2007, **3**:11–20.
59. Sakamaki M, Fujita S, Sato Y, Bemben MG, Abe T: **Blood pressure response to slow walking combined with KAATSU in the elderly.** *Int J Kaatsu Train Res* 2008, **4**:17–20.
60. Umbel JD, Hoffman RL, Dearth DJ, Chleboun GS, Manini TM, Clark BC: **Delayed-onset muscle soreness induced by low-load blood flow-restricted exercise.** *Eur J Appl Physiol* 2009, **107**:687–695.
61. Loenneke JP, Kearney ML, Thrower AD, Collins S, Pujol TJ: **The acute response of practical occlusion in the knee extensors.** *J Strength Cond Res* 2010, **24**:2831–2834.
62. Byrnes WC, Clarkson PM: **Delayed onset muscle soreness and training.** *Clin Sports Med* 1986, **5**:605–614.
63. Iversen E, Rostad V: **Low-load ischemic exercise-induced rhabdomyolysis.** *Clin J Sport Med* 2010, **20**:218–219.
64. Renzi CP, Tanaka H, Sugawara J: **Effects of leg blood flow restriction during walking on cardiovascular function.** *Med Sci Sports Exerc* 2010, **42**:726–732.
65. Mustoe T: **Understanding chronic wounds: a unifying hypothesis on their pathogenesis and implications for therapy.** *Am J Surg* 2004, **187**:655–705.
66. Wernbom M, Paulsen G, Nilsen TS, Hisdal J, Raastad T: **Contractile function and sarcolemmal permeability after acute low-load resistance exercise with blood flow restriction.** *Eur J Appl Physiol* 2008, **104**:813–819.
67. Goldfarb AH, Garten RS, Chee PD, Cho C, Reeves GV, Hollander DB, Thomas C, Aboudehen KS, Francois M, Kraemer RR: **Resistance exercise effects on blood glutathione status and plasma protein carbonyls: influence of partial vascular occlusion.** *Eur J Appl Physiol* 2008, **104**:813–819.
68. Loenneke JP, Wilson JM, Wilson GJ, Pujol TJ, Bemben MG: **Potential safety issues with blood flow restriction training.** *Scand J Med Sci Sports* 2011, **21**:510–518.
69. Nakajima T, Kurano M, Lida H, Takano H, Onuma H, Morita T, Meguro K, Sato Y, Nagata A: **Use and safety of KAATSU training: results of a national survey.** *Int J Kaatsu Train Res* 2006, **2**:5–13.
70. Cook SB, Brown KA, Deruisseau KC, Kanaley JA, Ploutz-Snyder LL: **Skeletal muscle adaptations following blood flow restricted training during 30 days of muscular unloading.** *J Appl Physiol* 2010, **2**:341–349.
71. Gopalakrishnan R, Genc KO, Rice AJ, Lee SM, Evans HJ, Maender CC, Ilaslan H, Cavanagh PR: **Muscle volume, strength, endurance, and exercise loads during 6-month missions in space.** *Aviat Space Environ Med* 2010, **81**:91–102.
72. LeBlanc A, Lin C, Shackelford L, Sinityn V, Evans H, Belichenko O, Schenkman B, Kozlovskaya I, Oganov V, Bakulin A, Hedrick T, Feeback D: **Muscle volume, MRI relaxation times (T2), and body composition after spaceflight.** *J Appl Physiol* 2000, **89**:2158–2164.

73. Trappe S, Costill D, Gallagher P, Creer A, Peters JR, Evans H, Riley DA, Fitts RH: **Exercise in space: human skeletal muscle after 6 months aboard the International Space Station.** *J Appl Physiol* 2009, **106**:1159–1168.
74. Sale D, Quinlan J, Marsh E, McComas AJ, Belanger AY: **Influence of joint position on ankle plantarflexion in humans.** *J Appl Physiol* 1982, **52**:1636–1642.
75. Nakajima T, Iida H, Kurano M, Takano H, Morita T, Meguro K, Sato Y, Yamazaki Y, Kawashima S, Ohshima H, Tachibana S, Ishii N, Abe T: **Hemodynamic responses to simulated weightlessness of 24-h head-down bed rest and KAATSU blood flow restriction.** *Eur J Appl Physiol* 2008, **104**:727–737.
76. Kubota N, Takano H, Tsutsumi T, Kurano M, Iida H, Yasuda T, Meguro K, Morita T, Sato Y, Kawashima S, Yamazaki Y, Ohshima H, Tachibana S, Ishii N, Abe T, Nakajima T: **Resistance exercise combined with KAATSU during simulated weightlessness.** *Int J Kaatsu Train Res* 2008, **4**:9–15.
77. Iida H, Kurano M, Takano H, Oonuma H, Imuta N, Kubota N, Morita T, Meguro K, Sato Y, Abe T, Yamazaki T, Nakajima T: **Can KAATSU be used for an orthostatic stress in astronauts? A case study.** *Int J Kaatsu Train Res* 2006, **2**:45–52.
78. Buckley JC Jr, Lane LD, Levine BD, Watenpaugh DE, Wright SJ, Moore WE, Gaffney FA, Blomqvist CG: **Orthostatic intolerance after spaceflight.** *J Appl Physiol* 1996, **81**:7–18.
79. Meck JV, Reyes CJ, Perez SA, Goldberger AL, Ziegler MG: **Marked exacerbation of orthostatic intolerance after long- vs. short-duration spaceflight in veteran astronauts.** *Psychosom Med* 2001, **63**:865–873.
80. Lee SM, Bennett BS, Hargens AR, Watenpaugh DE, Ballard RE, Murthy G, Ford SR, Fortney SM: **Upright exercise or supine lower body negative pressure exercise maintains exercise responses after bed rest.** *Med Sci Sports Exerc* 1997, **29**:892–900.
81. Fomina G, Kotovskaya A, Arbeille F, Pochuev V, Zhernavkov A, Ivanovskaya T: **Changes in hemodynamic and post-flights orthostatic tolerance of cosmonauts under application of the preventive device-thigh cuffs bracelets in short-term flights.** *J Gravit Physiol* 2004, **11**:229–230.
82. Herault S, Fomina G, Alferova I, Kotovskaya A, Poliakov V, Arbeille P: **Cardiac, arterial and venous adaptation to weightlessness during 6-month MIR spaceflights with and without thigh cuffs (bracelets).** *Eur J Appl Physiol* 2000, **81**:384–390.
83. Lee SM, Schneider SM, Boda WL, Watenpaugh DE, Macias BR, Meyer RS, Hargens AR: **Supine LBNP exercise maintains exercise capacity in male twins during 30-d bed rest.** *Med Sci Sports Exerc* 2007, **39**:1315–1326.
84. Perhonen MA, Franco F, Lane LD, Buckley JC, Blomqvist CG, Zerwekh JE, Peshock RM, Weatherall PT, Levine BD: **Cardiac atrophy after bed rest and spaceflight.** *J Appl Physiol* 2001, **91**:645–653.
85. Levine BD, Zuckerman JH, Pawelczyk JA: **Cardiac atrophy after bed-rest deconditioning: a nonneural mechanism for orthostatic intolerance.** *Circulation* 1997, **96**:517–525.
86. Shibata S, Perhonen M, Levine BD: **Supine cycling plus volume loading prevent cardiovascular deconditioning during bed rest.** *J Appl Physiol* 2010, **108**:1177–1186.
87. Folkow B: **Structure and function of the arteries in hypertension.** *Am Heart J* 1987, **114**:938–948.
88. Taday EC, Meck JV, Nyhan D, Shoukas AA, Berkowitz DE: **Microgravity-induced changes in aortic stiffness and their role in orthostatic intolerance.** *J Appl Physiol* 2007, **102**:853–858.
89. Hesse C, Siedler H, Luntz SP, Arendt BM, Goerlich R, Fricker R, Heer M, Haefeli WE: **Modulation of endothelial and smooth muscle function by bed rest and hypoenergetic, low-fat nutrition.** *J Appl Physiol* 2005, **99**:2196–2203.
90. Seals DR: **Habitual exercise and the age-associated decline in large artery compliance.** *Exerc Sport Sci Rev* 2003, **31**:68–72.
91. Tanaka H, Dinunno FA, Monahan KD, Clevenger CM, DeSouza CA, Seals DR: **Aging, habitual exercise, and dynamic arterial compliance.** *Circulation* 2000, **102**:1270–1275.
92. Kim SJ, Sherk VD, Bemben MG, Bemben DA: **Effects of short term, low intensity resistance training with vascular restriction on arterial compliance in untrained young men.** *Int J Kaatsu Train Res* 2009, **5**:1–8.
93. Ozaki H, Miyachi M, Nakajima T, Abe T: **Effects of 10 weeks walk training with leg blood flow reduction on carotid arterial compliance and muscle size in the elderly adults.** *Angiology* 2011, **62**:81–86.
94. Celermajer DS, Sorensen KE, Gooch VM, Spiegelhalter DJ, Miller OI, Sullivan ID, Lloyd JK, Deanfield JE: **Non-invasive detection of endothelial dysfunction in children and adults at risk of atherosclerosis.** *Lancet* 1992, **340**:1111–1115.
95. Anderson TJ, Uehata A, Gerhard MD, Meredith IT, Knab S, Delagrangre D, Lieberman EH, Ganz P, Creager MA, Yeung AC, Selwyn AP: **Close relation of endothelial function in the human coronary and peripheral circulations.** *J Am Coll Cardiol* 1995, **26**:1235–1241.
96. Schroeder S, Enderle MD, Ossen R, Meisner C, Baumbach A, Pfohl M, Herdeg C, Oberhoff M, Haering HU, Karsch KR: **Noninvasive determination of endothelium-mediated vasodilation as a screening test for coronary artery disease: pilot study to assess the predictive value in comparison with angina pectoris, exercise electrocardiography, and myocardial perfusion imaging.** *Am Heart J* 1999, **138**:731–739.
97. Monahan KD, Tanaka H, Dinunno FA, Seals DR: **Central arterial compliance is associated with age- and habitual exercise-related differences in cardiovagal baroreflex sensitivity.** *Circulation* 2001, **104**:1627–1632.
98. Grigoriev AI, Bugrov SA, Bogomolov VV, Egorov AD, Kozlovskaya IB, Pestov ID, Polyakov VV, Tarasov IK: **Preliminary medical results of the Mir year-long mission.** *Acta Astronaut* 1991, **23**:1–8.
99. Biering-Sorensen F, Bohr HH, Schaadt OP: **Longitudinal study of bone mineral content in the lumbar spine, the forearm and the lower extremities after spinal cord injury.** *Eur J Clin Invest* 1990, **20**:330–335.
100. Lang T, LeBlanc A, Evans H, Lu Y, Genant H, Yu A: **Cortical and trabecular bone mineral loss from the spine and hip in long-duration spaceflight.** *J Bone Miner Res* 2004, **19**:1006–1012.
101. LeBlanc A, Schneider V, Shackelford L, West S, Oganov V, Bakulin A, Voronin L: **Bone mineral and lean tissue loss after long duration space flight.** *J Musculoskelet Neuronal Interact* 2000, **1**:157–160.
102. Bemben DA, Palmer IJ, Abe T, Sato Y, Bemben MG: **Effects of a single bout of low intensity KAATSU resistance training on markers of bone turnover in young men.** *Int J Kaatsu Train Res* 2007, **3**:21–26.
103. Fritton SP, Weinbaum S: **Fluid and solute transport in bone: flow-induced mechanotransduction.** *Annu Rev Fluid Mech* 2009, **41**:347–374.
104. Loenneke JP, Young KC, Fahs CA, Rossow LM, Bemben DA, Bemben MG: **Blood flow restriction: rationale for improving bone.** *Med Hypotheses* 2012, **78**:523–527.
105. Hitt D, Garriott O, Kerwin J: *Homesteading in Space: The Skylab Story.* Lincoln and London: University of Nebraska Press; 2008.
106. Walker DK, Dickinson JM, Timmerman KL, Drummond MJ, Reidy PT, Fry CS, Gundermann DM, Rasmussen BB: **Exercise, amino acids, and aging in the control of human muscle protein synthesis.** *Med Sci Sports Exerc* 2011, **43**:2249–2258.
107. Ozaki H, Sakamaki M, Yasuda T, Fujita S, Ogasawara R, Sugaya M, Nakajima T, Abe T: **Increases in thigh muscle volume and strength by walk training with leg blood flow reduction in older participants.** *J Gerontol A Biol Sci Med Sci* 2011, **66**:257–263.
108. Abe T, Sakamaki M, Fujita S, Ozaki H, Sugaya M, Sato Y, Nakajima T: **Effects of low-intensity walk training with restricted leg blood flow on muscle strength and aerobic capacity in older adults.** *J Geriatr Phys Ther* 2010, **33**:34–40.
109. Gualano B, Neves M Jr, Lima FR, Pinto AL, Laurentino G, Borges C, Baptista L, Artioli GG, Aoki MS, Moricot A, Lancha AH Jr, Bonfá E, Ugrinowitsch C: **Resistance training with vascular occlusion in inclusion body myositis: a case study.** *Med Sci Sports Exerc* 2010, **42**:250–254.
110. Nakajima T, Kurano M, Sakagami F, Iida H, Fukumura K, Fukuda T, Takano H, Madarame H, Yasuda T, Nagata T, Sato Y, Yamasoba T, Morita T: **Effects of low intensity KAATSU resistance training on skeletal muscle size/strength and endurance capacity in patients with ischemic heart disease.** *Int J Kaatsu Train Res* 2010, **6**:1–7.
111. Sumide T, Sakuraba K, Sawaki K, Ohmura H, Tamura Y: **Effect of resistance exercise training combined with relatively low vascular occlusion.** *J Sci Med Sport* 2009, **12**:107–112.
112. Yokokawa Y, Hongo M, Urayama H, Nishimura T, Kai I: **Effects of low-intensity resistance exercise with vascular occlusion on physical function in healthy elderly people.** *Biosci Trends* 2008, **2**:117–123.
113. Kubo K, Komuro T, Ishiguro N, Tsunoda N, Sato Y, Ishii N, Kanehisa H, Fukunaga T: **Effects of low-load resistance training with vascular occlusion on the mechanical properties of muscle and tendon.** *J Appl Biomech* 2006, **22**:112–119.
114. Abe T, Beekley MD, Hinata S, Koizumi K, Sato Y: **Day-to-day changes in muscle strength and MRI-measured skeletal muscle size during 7 days of KAATSU resistance training.** *Int J Kaatsu Train Res* 2005, **1**:71–76.

115. Ishii N, Madarame H, Odagiri K, Naganuma M, Shinoda K: **Circuit training without external load induces hypertrophy in lower-limb muscles when combined with moderate venous occlusion.** *Int J Kaatsu Train Res* 2005, **1**:24–28.
116. Takarada Y, Sato Y, Ishii N: **Effects of resistance exercise combined with vascular occlusion on muscle function in athletes.** *Eur J Appl Physiol* 2002, **86**:308–314.
117. Shinohara M, Kouzaki M, Yoshihisa T, Fukunaga T: **Efficacy of tourniquet ischemia for strength training with low resistance.** *Eur J Appl Physiol Occup Physiol* 1998, **77**:189–191.
118. Moore DR, Burgomaster KA, Schofield LM, Gibala MJ, Sale DG, Phillips SM: **Neuromuscular adaptations in human muscle following low intensity resistance training with vascular occlusion.** *Eur J Appl Physiol* 2004, **92**:399–406.
119. Takarada Y, Takazawa H, Sato Y, Takebayashi S, Tanaka Y, Ishii N: **Effects of resistance exercise combined with moderate vascular occlusion on muscular function in humans.** *J Appl Physiol* 2000, **88**:2097–2106.
120. Sato Y, Yashitomi A, Abe T: **Acute growth hormone response to low-intensity KAATSU resistance exercise: comparison between arm and leg.** *Int J Kaatsu Train Res* 2005, **1**:45–50.
121. Tanimoto M, Madarame H, Ishii N: **Muscle oxygenation and plasma growth hormone concentration during and after resistance exercise: comparison between "KAATSU" and other types of regimen.** *Int J Kaatsu Train Res* 2005, **1**:51–56.
122. Madarame H, Kurano M, Takano H, Iida H, Sato Y, Ohshima H, Abe T, Ishii N, Morita T, Nakajima T: **Effects of low-intensity resistance exercise with blood flow restriction on coagulation system in healthy subjects.** *Clin Physiol Funct Imaging* 2010, **30**:210–213.
123. Iida H, Kurano M, Takano H, Kubota N, Morita T, Meguro K, Sato Y, Abe T, Yamazaki Y, Uno K, Takenaka K, Hirose K, Nakajima T: **Hemodynamic and neurohumoral responses to the restriction of femoral blood flow by KAATSU in healthy subjects.** *Eur J Appl Physiol* 2007, **100**:275–285.
124. Kubota A, Sakuraba K, Sawaki K, Sumide T, Tamura Y: **Prevention of disuse muscular weakness by restriction of blood flow.** *Med Sci Sports Exerc* 2008, **40**:529–534.

doi:10.1186/2046-7648-1-12

Cite this article as: Hackney *et al.*: Blood flow-restricted exercise in space. *Extreme Physiology & Medicine* 2012 **1**:12.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

